

# Linear robustness analysis of non-linear biomolecular networks with kinetic perturbations

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## Robustness analysis



#### Informal definition of robustness

Robustness is a property that allows a system to maintain its function in the presence of internal and external perturbations.

H. Kitano, 2004

- System / Model:  $\dot{x} = Sv(x)$ 
  - $x \in \mathbb{R}^n$ : amounts of signalling molecules
  - $v(x) \in \mathbb{R}^m$ : reaction rates
  - $S \in \mathbb{R}^{n \times m}$ : stoichiometric matrix
- Perturbations: kinetic perturbations (changes in reaction kinetics v)
- Function: Qualitative dynamical behaviour (stability, oscillations, ...)



- Non-robust perturbation = any deviation from the nominal point for which the system looses functionality
- Robustness measure = smallest norm of non-robust perturbations

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## Robustness analysis for model evaluation

- Biological systems are robust ↔ models should be robust!
- Find parts of the system which need to be modelled carefully
- Evaluate effects of uncertainty on model predictions

## Robustness analysis for system understanding

- Understand mechanisms which confer robustness to biological systems
- Detect fragile points, e.g. for medical intervention
- Guide system design in bioengineering and synthetic biology



## Outline



- Theory of kinetic perturbations
- Pobustness analysis with kinetic perturbations
- 3 Adaptation with kinetic perturbations

## Outline

## Theory of kinetic perturbations

- Definition of kinetic perturbations
- Mapping kinetic perturbations to parameter variations

2 Robustness analysis with kinetic perturbations

3 Adaptation with kinetic perturbations

## Kinetic perturbations - the basic idea



## Kinetic perturbations = reaction rate perturbations

- nominal rates v(x) to perturbed rates  $\tilde{v}(x)$
- $\Rightarrow$  nominal model  $\dot{x} = Sv(x)$  to perturbed model  $\dot{x} = S\tilde{v}(x)$ 
  - Consider nominal steady state  $x_0$ :  $Sv(x_0) = 0$
  - Kinetic perturbation, if  $\tilde{v}(x_0) = v(x_0)$
  - Reaction rates in steady state unperturbed, but reaction rate slopes may vary:

     <sup>x</sup>(x) increased slope





#### Definition

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The network  $\dot{x} = S\tilde{v}(x)$  is said to be a *kinetic perturbation* of the nominal network  $\dot{x} = Sv(x)$  at  $x_0 \in \mathbb{R}^n$  with  $Sv(x_0) = 0$ , if  $\tilde{v}(x_0) = v(x_0) =: v_0$ .

- Consider the reaction rate Jacobian  $V = \frac{\partial V}{\partial x}$
- For the perturbed network, we have

$$\tilde{V}(x_0) = \frac{\partial \tilde{V}}{\partial x}(x_0) = V(x_0) + \bar{\Delta}$$

- Local effects of the kinetic perturbation are determined by  $\bar{\Delta}$ 
  - Scaled perturbation  $\Delta_{i\ell} = \frac{x_{0,\ell}}{v_{0,i}} \bar{\Delta}_{i\ell}$
  - $\|\Delta\|$  is a measure for the perturbation size
- Steady state concentration x<sub>0</sub> and flux v<sub>0</sub> are unperturbed
- Steady state reaction slopes  $V(x_0)$  are perturbed by  $\bar{\Delta}$

# Mapping kinetic perturbations to parameter changes

- Considering generalised mass action (GMA) networks
- GMA reaction rate:  $v_i(x) = k_i \prod_{\ell=1}^n x_{\ell}^{\alpha_{i\ell}}$ 
  - $\alpha_{i\ell} > 0$ :  $x_{\ell}$  is a substrate for / activates  $v_i$
  - $\alpha_{i\ell} < 0$ :  $x_{\ell}$  inhibits  $v_i$
- Non-integer kinetic orders α<sub>iℓ</sub>
  - Effect of constrained diffusion (fractal reaction kinetics)
  - Result of model simplifications (as in S-systems, Savageau et al.)
- Perturbed reaction rate  $\tilde{v}_i(x) = \tilde{k}_i \prod_{\ell=1}^n x_\ell^{\tilde{\alpha}_{i\ell}}$



Are kinetic perturbations biochemically plausible?

Yes, because kinetic perturbations map to parameter variations in reasonable model classes for intracellular networks such as

- GMA networks
- enzymatic networks (Michaelis-Menten or Hill reaction kinetics)
- genetic networks (sigmoidal activation & inhibition functions)

#### **Biochemical interpretation**

- Positive Δ<sub>i</sub>: more cooperativity in reaction *i* with respect to species *l*
- Negative Δ<sub>il</sub>: increase saturation in reaction *i* with respect to species l

## Outline



## Robustness analysis with kinetic perturbations

- Kinetic perturbations affect dynamical properties
- Robustness analysis via the structured singular value
- Example: Oscillations in the MAPK cascade

3 Adaptation with kinetic perturbations



# Inducing bifurcations by kinetic perturbations

Example "network"

$$\xrightarrow{\psi} V_1 = \frac{kx^2}{1+Mx^2} \qquad \dot{x} = V_1 - V_2$$

$$\xrightarrow{\psi} V_1 \Rightarrow \qquad V_2 = x$$



Loss of bistability by a kinetic perturbation

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# Inducing bifurcations by kinetic perturbations

Example "network"

$$\xrightarrow{\downarrow} V_1 = \frac{kx^2}{1+Mx^2} \qquad \dot{x} = V_1 - V_2$$

$$\xrightarrow{\psi} V_1 \rightarrow X \qquad \xrightarrow{V_2} \qquad V_2 = x$$



Loss of bistability by a kinetic perturbation

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# Inducing bifurcations by kinetic perturbations

Example "network"

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Loss of bistability by a kinetic perturbation



# Robustness with respect to kinetic perturbations



#### Perturbed Jacobian

$$\tilde{A}(\Delta) = A + S(\operatorname{diag} v_0) \Delta(\operatorname{diag} x_0)^{-1}$$

## Robustness radius for kinetic perturbations

• Robustness measure = size of smallest perturbation such that  $\tilde{A}(\Delta)$  has an eigenvalue on the imaginary axis

$$\psi = \inf\{\|\Delta\| \mid \det(j\omega I - \tilde{A}(\Delta)) = 0\}$$

• Can be formulated as structured singular value ( $\mu$ ) problem with  $G(j\omega) = (\operatorname{diag} x_0)^{-1} (j\omega I - A)^{-1} S(\operatorname{diag} v_0)$ 

Result: 
$$\psi = \left(\sup_{\omega} \mu_{\Delta} G(j\omega)\right)^{-1}$$

Robustness problem can be solved with standard  $\mu$  analysis tools.



# Robustness radius for scalar perturbations



• Perturbation of a single interaction: Species  $\ell$  on reaction *i* 

$$\bar{\Delta} = \begin{pmatrix} 0 \\ \vdots \\ 1 \\ \vdots \\ 0 \end{pmatrix} \bar{\Delta}_{i\ell} (0 \dots 1 \dots 0) = \boldsymbol{e}_i \bar{\Delta}_{i\ell} \boldsymbol{e}_{\ell}^{\mathrm{T}}$$

• Perturbed Jacobian  $\tilde{A}(\Delta_{i\ell}) = A + Se_i v_{0,i} \Delta_{i\ell} x_{0,\ell}^{-1} e_{\ell}^T$ 



- $\mu$ -problem with  $G_{\ell i}(j\omega) = e_{\ell}^{\mathrm{T}} x_{0,\ell}^{-1} (j\omega I A)^{-1} S v_{0,i} e_{i}$
- Explicit formula for the robustness radius
- Allows to detect fragile interactions in the network

## Robustness analysis in the vector case



- Perturbation of several interactions either
  - from one species  $\ell$  to several reactions  $E_i v$  or
  - from several species  $E_{\ell}x$  to one reaction *i*.

$$\bar{\Delta} = E_i \bar{\Delta}_{i\ell} e_\ell^{\mathrm{T}}$$

• Perturbed Jacobian  $\tilde{A}(\Delta_{i\ell}) = A + SE_i \operatorname{diag}(E_i v_0) \Delta_{i\ell} x_{0,\ell}^{-1} e_{\ell}^T$ 

## Robustness analysis in the vector case

- Results depend on the norm that is chosen:
  - 1-norm or  $\infty$ -norm: robustness radius from a linear program
  - 2-norm: explicit formula for robustness radius
- Also requires to compute a supremum over  $\omega$ .
- Non-robust perturbation is the solution of an affine equation  $M\bar{\Delta} = b$ .



## The MAPK cascade



Medium scale network with 22 species and 30 (irreversible) reactions

Where to perturb this network in order to get oscillations?

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- Checking all 660 network interactions for a scalar kinetic perturbation to induce oscillations
- Found 8 fragile interactions (|Δ\*| < 1), e.g.:</p>



- Negative feedback from MAPKpp to MAPKK phosphorylation
- Positive feedback from MAPKpp to catalysis of MAPK / MAPKKpp complex
- Saturated catalysis of MAPKpp / Phosphatase complex

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Effect of a kinetic perturbation on the dynamics





Critical perturbation  $\Delta^* = -0.76$ 



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Effect of a kinetic perturbation on the dynamics





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Effect of a kinetic perturbation on the dynamics





Critical perturbation  $\Delta^* = -0.76$ 



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## Outline



## Theory of kinetic perturbations

2 Robustness analysis with kinetic perturbations

- Adaptation with kinetic perturbations
  - Properties of biomolecular adaptation
  - Achieving adaptation with kinetic perturbations
  - Example: Adaptation in the MAPK cascade

## Definition of adaptation



## 1. Adaptation

A dynamical system  $u \mapsto y$  is said to adapt, if persistent changes in udo lead to transient, but not persistent changes in y.



Local adaptation = consider only infinitesimally small changes in u

# 2. Local adaptation without transient response The system $u \mapsto y$ is said to perfectly adapt locally at $(u_0, y_0)$ , if

$$\lim_{u_s\to 0}\frac{\lim_{t\to\infty}(y(t,u_0+u_sh(t))-y_0)}{u_s}=0.$$



## Adaptation and robustness



Adaptation to a slowly changing perturbation

Robustness of steady state concentration to this perturbation



## Slowly changing perturbations in biology

- Protein amounts due to stochastic gene expression
- Environmental conditions such as light, temperature, or nutrient availability



## Conditions for adaptation

### 1. Linear approximation at steady state

$$dx = Adx + Bdu$$
 with:  $A = S \frac{\partial V}{\partial x}(x_0, u_0)$   
 $dy = Cdx,$   $B = S \frac{\partial V}{\partial u}(x_0, u_0)$ 

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2. Tranformation to frequency domain

$$dy(s) = C(sI - A)^{-1}Bdu(s) = G(s)du(s).$$

3. Necessary and sufficient condition for local adaptation By the final value theorem of the Laplace transformation:

local adaptation 
$$\Leftrightarrow$$
  $G(0) = 0 \stackrel{A \text{ invertible}}{\Leftrightarrow} \det \begin{pmatrix} A & B \\ C & 0 \end{pmatrix} = 0$ 

Last condition is affected by kinetic perturbations!

# Achieving adaptation with kinetic perturbations



## Effect of kinetic perturbation on adaptation condition

• Perturbation of Jacobian due to a kinetic perturbation  $\ell \rightarrow i$ :

$$\tilde{A} = A + Se_i v_{0,i} \Delta_{i\ell} (x_{0,\ell})^{-1} e_{\ell}^{\mathrm{T}}$$

Adaptation as goal:

$$\det\begin{pmatrix} \tilde{A} & B\\ C & 0 \end{pmatrix} = \det\begin{pmatrix} A + Se_i v_{0,i} \Delta_{i\ell}(x_{0,\ell})^{-1} e_{\ell}^{\mathrm{T}} & B\\ C & 0 \end{pmatrix} = 0.$$

Compute a critical kinetic perturbation

$$\Delta_{i\ell}^{*} = (-e_{j}^{\mathrm{T}}M_{out}M_{0}^{-1}M_{in}e_{\ell})^{-1}\frac{x_{0,\ell}}{v_{0,i}}$$

with

$$M_0 = \begin{pmatrix} A & B \\ C & 0 \end{pmatrix} \qquad M_{in} = \begin{pmatrix} I \\ 0 \end{pmatrix} S \qquad M_{out} = \begin{pmatrix} I & 0 \end{pmatrix}$$



# Example: Adaptation in the MAPK cascade



## Goal: achieve adaptation of nuclear MAPKpp



- Huang-Ferrell MAPK cascade model as before
- Added an input by controlling E1 production

#### Result of analysis with kinetic perturbations

- No adaptation with transient response possible through kinetic perturbations
- Network lacks a feedback or feedforward loop

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### Model feedback extension



- Added a protein genetically regulated by MAPK
- Feedback by formation of inhibitory complex

#### Candidate interactions for adaptation

Found five candidate interactions in newly added feedback loop for adaptation via kinetic perturbations.



## Response of original and perturbed networks





- Adaptation achieved even for large stimulus changes.
- Non-linear dynamics depend on which interaction is targeted.

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### Kinetic perturbations

- Perturbation class suited for biochemical network analysis
- Kinetic perturbations = reaction rate slope changes at steady state
- Biochemically plausible for GMA / enzymatic / genetic networks

## Robustness analysis with kinetic perturbations

- Efficient robustness analysis via µ–analysis methods
- Results indicate fragile network interactions
- Non-robust perturbations are explicitly computed

## Adaptation with kinetic perturbations

- Find network modifications to achieve perfect local adaptation
- Steady state is kept unperturbed intrinsically
- Adaptation even to large stimulus changes in many cases